

This article was downloaded by:

On: 23 January 2011

Access details: *Access Details: Free Access*

Publisher *Taylor & Francis*

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



## Journal of Carbohydrate Chemistry

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713617200>

### Synthesis of N-Hetarylthiourea Derivatives of Carbohydrates

José Fuentes Mota; Fernández José M. García; Carmen Ortiz Mellet; Pradera Adrián M. Angeles; Tomasa Cuevas Lorite

**To cite this Article** Mota, José Fuentes , García, Fernández José M. , Mellet, Carmen Ortiz , Angeles, Pradera Adrián M. and Lorite, Tomasa Cuevas(1990) 'Synthesis of N-Hetarylthiourea Derivatives of Carbohydrates', *Journal of Carbohydrate Chemistry*, 9: 6, 837 – 851

**To link to this Article:** DOI: 10.1080/07328309008543878

**URL:** <http://dx.doi.org/10.1080/07328309008543878>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

SYNTHESIS OF N-HETARYLTHIOUREA DERIVATIVES OF CARBOHYDRATES<sup>1</sup>

José Fuentes Mota , José M. García Fernández, Carmen Ortiz Mellet,  
M. Angeles Pradera Adrián and Tomasa Cuevas Lorite

Departamento de Química Orgánica, Facultad de Química,  
Universidad de Sevilla, Apartado 553, 41071 Sevilla, Spain.

Received January 30, 1990 - Final Form May 31, 1990

ABSTRACT

The syntheses of N-hetaryl(thiazole-2-yl, 2-thiazoline-2-yl, 4,4-diphenyloxazoline-2-yl, *cis*-3a,4,5,6,7,7a-hexahydrobenzoxazole-2-yl)-N'-(2,3,4,6-tetra-O-acetyl-β-D-glucopyranosyl)thioureas (1a-1d), N-hetaryl(2-thiazoline-2-yl, 4,4-diphenyloxazoline-2-yl)-N'-(2,3,4,6-tetra-O-benzoyl-β-D-galactopyranosyl)thioureas (2b, 2c) and 1,3,4,6-tetra-O-acetyl-2-deoxy-2-[3-(4',4'-diphenyl-2'-yl)thioureido]-β-D-glucopyranose (3c) are described. The structures and conformational properties of prepared compounds are based on analytical and spectroscopic (UV, IR, NMR and MS) data.

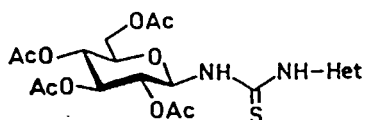
INTRODUCTION

The glycosylthioureas are compounds of pharmaceutical interest and they have been widely used in the syntheses of N-nucleosides and

glycosylaminoheterocycles.<sup>2,3</sup> As far as we are aware reports of hetarylthiourea carbohydrate derivatives are limited and generally report on the syntheses of pyridine derivatives<sup>2,4</sup> and compounds with two sugar moieties.<sup>2,5</sup> We now describe the preparation of the N-glycosyl-N'-hetarylthioureas 1a-1d, 2b, 2c and the 2-deoxy-2-thioureidogluco-Syl derivative 3c.

## RESULTS AND DISCUSSION

The acylated glycosylthioureas 1a-1d, 2b and 2c were prepared by reaction of the corresponding per-O-acylglycosyl isothiocyanates and hetarylamines. Compound 1d was isolated as a pair of diastereomers

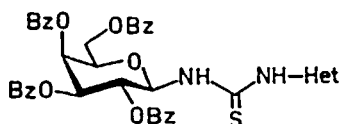


1a - 1d

Het =



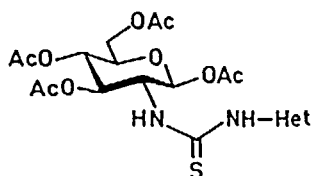
a



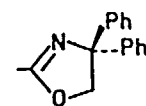
2b, 2c



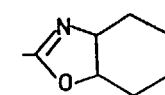
b



3c



c



d

Table 1.  $^1\text{H}$  NMR chemical shifts ( $\delta$ , ppm) of compounds 1a-1d, 2b, 2c and 3c in  $\text{CDCl}_3$ 

Comp.	Sugar moiety						NH	N'H	Heterocycle		
	H-1	H-2	H-3	H-4	H-5	H-6			H-6'	H-4(4',3a)	H-5(5',7a)
<u>1a</u>	5.83t	5.22t	5.36t	5.13t	3.90ddd	4.30dd	4.14dd	bs <sup>a</sup>	bs <sup>a</sup>	7.39d(1H)	6.88d(1H)
<u>1b</u>	5.77dd	5.17t	5.32t	5.11t	3.85ddd	4.28dd	4.17dd	13.08d	8.20bs	4.74m(2H)	3.17t(2H)
<u>1c</u> <sup>b</sup>	5.75t	5.18-4.92	5.36t	5.18-4.92	3.88ddd	4.31dd	4.16dd	11.90bs	7.05bd	--	4.89s(2H)
<u>1c</u>	5.84bt	5.05t	5.32t	4.94t	←-----	4.20-3.85m	----->	11.95bs	7.05bd	--	4.94s(2H)
<u>1d</u>	5.78t	5.18t	5.33t	5.12t	3.86m	4.33dd	4.15dd	6.43bs	12.30d	4.48m(1H)	4.92m(1H)
<u>2b</u>	5.76t	5.16t	5.32t	5.11t	3.86m	4.28dd	4.15dd	6.43bs	12.32d	4.48m(1H)	4.92m(1H)
<u>2c</u> <sup>b</sup>	6.14dd	5.91t	5.76t	6.03d	←-----	4.65-4.35m	----->	13.36d	8.20bs	4.71m(2H)	3.08t(2H)
<u>3c</u> <sup>b</sup>	←-----	6.20-5.66m	←-----	←-----	4.60-4.35m	4.65dd	4.60-4.35m	11.95bs	8.20-7.20	--	4.85m(2H)
	5.84d	5.10q	5.31t	5.20t	3.86m	4.30dd	4.13dd	6.61d	11.85bs	--	4.86bs

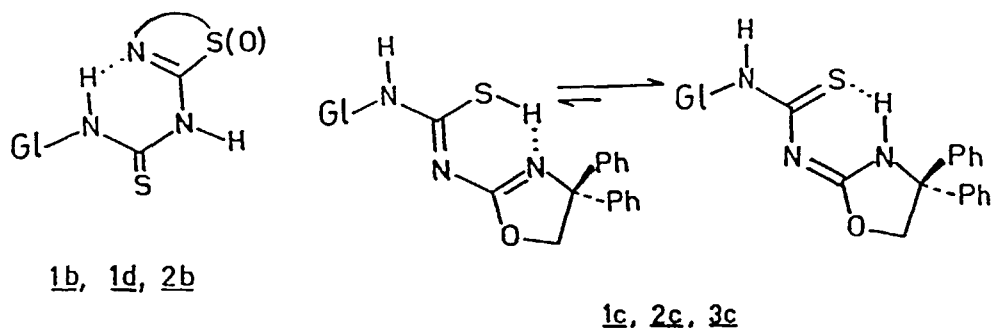
a. Very broad peak indistinguishable from the base line. b. Data for major tautomer. c. In  $(\text{CD}_3)_2\text{SO}$  at 130°C. d. Pair of diastereomers.

Table 2.  $^1\text{H}$  NMR coupling constants (Hz) of compounds 1a-1d, 2b, 2c and 3c in  $\text{CDCl}_3$ 

Compound	Sugar moiety							Heterocycle	
	$J_{1,2}$	$J_{2,3}$	$J_{3,4}$	$J_{4,5}$	$J_{5,6}$	$J_{5,6'}$	$J_{6,6'}$	$J_{1(2),\text{NH}}$	$J_{4,5}$
<u>1a</u>	9.3	9.3	9.3	9.3	4.2	1.7	12.3	9.3	3.7
<u>1b</u>	9.7	9.7	9.7	9.7	4.5	2.3	12.5	8.6	6.8
<u>1c</u>	9.3	9.3	9.3	9.3	4.4	1.8	10.6	9.3	-
<u>1d</u>	9.0	9.0	9.0	9.0	4.8	2.3	13.2	9.0	-
<u>2b</u>	9.3	9.3	3.4	0.0	-	-	-	7.9	7.0
<u>2c</u>	-	-	-	-	5.6	-	11.0	-	-
<u>3c</u>	8.7	8.7	8.7	8.7	5.0	2.5	12.5	8.7	-

since the starting amine was a racemic mixture.<sup>8</sup> In the case of galactopyranosyl compounds (2b and 2c), the benzoyl group was used as protecting group since the corresponding acetyl derivatives could not be isolated with analytical purity. Compound 3c was prepared by reaction of 1,3,4,6-tetra-0-acetyl-2-deoxy-2-isothiocyanato- $\beta$ -D-glucopyranose and 2-amino-4,4-diphenyl-2-oxazoline.

The structures of compounds 1a-1d, 2b, 2c and 3c were based on analytical, UV, IR and  $^1\text{H}$  (Tables 1, 2) and  $^{13}\text{C}$  NMR (Table 3) data. The product 1a had  $\lambda_{\text{max}}$  at 253 nm, 1b at 274 nm, 1c, 1d and 3c at 230-234 nm, characteristic of thiazole,<sup>9</sup> 2-thiazoline<sup>5</sup> and aminooxazoline<sup>8</sup> rings respectively. The UV absorptions of heterocyclic moieties of 2b-2c overlapped those of the benzoyl groups. The shift at lower wavenumber of NH ( $\Delta\nu = 250 \text{ cm}^{-1}$ ) and C=N ( $\Delta\nu 20-40 \text{ cm}^{-1}$ ), with respect to other non-bonded thiazolines, 2-amino-2-thiazolines and 2-amino-2-oxazolines,<sup>10</sup> indicates that a strong intramolecular hydrogen bond is



Gl = SUGAR MOIETY

Scheme 1

present in compounds 1b-1d, 2b-2c and 3c (Scheme 1). The wavenumber of  $\nu_{\text{NH}}$  and  $\nu_{\text{CN}}$  in 1b and 2b are coincident with those reported for N-(2,3,4,6-tetra-O-acetyl- $\beta$ -D-glucopyranosyl)-N'-(4-chlorophenyl)-N-(2-thiazolin-2-yl)thiourea 4,<sup>5</sup> where the NH is intramolecularly bonded. Dilution studies in carbon tetrachloride solutions<sup>11</sup> performed on 1b, 2b and 1d confirmed the intramolecular bonds. The spectra showed a broad absorption with maxima at 3098-3030  $\text{cm}^{-1}$  whose relative absorbance with respect to  $\nu_{\text{CH}}$  (alkyl) was invariable with dilution ( $1 \cdot 10^{-2}$  -  $0.25 \cdot 10^{-2}$  molar). The  $\delta$  values of NH in 1b and 2b (13.08 and 13.36 ppm) also agree with the presence of these hydrogen bonds (Scheme 1), in agreement with that previously described for related compounds.<sup>5</sup>

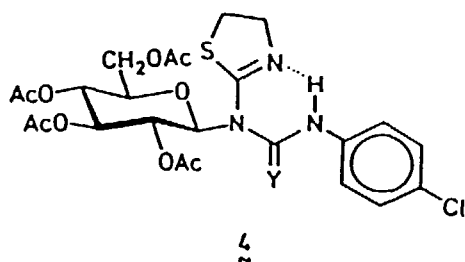


Table 3.  $^{13}\text{C}$  NMR chemical shifts ( $\delta$ , ppm) for the sugar moieties of compounds 1a-1d, 2b, 2c and 3c in  $\text{CDCl}_3$ .

	C-1	C-2	C-3	C-4	C-5	C-6
<u>1a</u>	82.7	70.2	73.0	68.1	73.5	61.6
<u>1b</u>	82.6	69.8	73.2 <sup>a</sup>	68.1	73.3 <sup>a</sup>	61.6
<u>1c</u> <sup>b</sup>	82.0	70.1	73.1 <sup>a</sup>	68.1	73.3 <sup>a</sup>	61.5
<u>1d</u>	82.6	70.1	73.3 <sup>a</sup>	68.1	73.5 <sup>a</sup>	61.6
	83.1	70.0	72.7 <sup>a</sup>	68.4	73.1 <sup>a</sup>	61.9
<u>2b</u>	83.2	68.5 <sup>a</sup>	72.1	68.3 <sup>a</sup>	73.0	62.1
<u>2c</u> <sup>b</sup>	82.7	69.2	71.8	68.2	72.7	61.9
<u>3c</u>	92.0	55.9	72.6 <sup>a</sup>	67.6	72.5 <sup>a</sup>	61.4

a. Assignments may be interchanged. b. Signals for the major compound.

The  $^1\text{H}$  NMR spectra of 1c, 2c and 3c indicated that these compounds in methyl sulphoxide solutions exist as the tautomeric equilibrium mixture shown in Scheme 1. The  $^1\text{H}$  resonances were pairs of signals with similar  $J$  values which coalesced to single signals on heating at 130 °C. The  $\delta$  values of NH (SH) are indicative of hydrogen bonds. The magnitudes of  $J_{1,\text{NH}}$  ( $J_{2,\text{N}'\text{H}}$  in 3c) are 8.7-9.3 Hz, showing antiperiplanar position<sup>12</sup> for N'H(NH) and H-1(H-2) and ruling out other possible hydrogen bonds. The assignments of  $^{13}\text{C}$ -resonances are based on APT spectra<sup>13</sup> and literature data for glycosyl<sup>3,5,7,14,15</sup> and heterocyclic<sup>5,12</sup> related compounds.

The N-(cis-3a,4,5,6,7,7a-hexahydrobenzoxazole-2-yl)-N'-(2,3,4,6-tetra-O-acetyl- $\beta$ -D-glucopyranosyl)thiourea (1d) was obtained by reaction

of the corresponding isothiocyanate and the racemic amine. According to the  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra **1d** was isolated as a pair of diastereomers. The  $\delta$  values of N'H (12.30d and 12.32d) are indicative of the hydrogen bonds shown in Scheme 1.

The  $^3J_{\text{H,H}}$  values for **1a-1d**, **2b** and **3c** showed that the  $^4C_1(\underline{D})$  conformation (sugar ring) preponderates in solutions in chloroform and methyl sulphoxide. Generally 1,3-disubstituted alkylthioureas exist in solution in conformational equilibrium (C-N bonds) between the (Z,E), (Z,Z) and (E,Z) forms.<sup>16,17</sup> In compounds **1b**, **1c** and **2b** the (Z,E) conformation is fixed by an internal hydrogen bond (Scheme 1). The  $J_{1,\text{NH}}$  and  $\delta$  H-1 values agree with those previously described for (Z,E) conformations of alkylthioureas.<sup>18</sup>

The mass spectra of per-O-acetylglucopyranosylthioureas (**1a-1d**) show as the main fragmentation pathway the cleavage of the glycosidic bond<sup>19</sup> ( $m/z$  331) and formation of the fragments  $m/z$  271, 211, 169, 127, 109, characteristic of per-O-acetyl sugar derivatives.<sup>20-21</sup> Other significant peaks are assigned to hetaryl isothiocyanates<sup>19</sup> and hetaryl amines. Additionally compound **1a** showed the described fragments of 2-aminothiazole<sup>22</sup> and **1b** the fragments of 2-aminothiazoline<sup>22</sup> (see Experimental). The compound **1c** shows an ion at  $m/z$  165(23%) which is assigned to the fluorenyl cation.

The mass spectra of **2b** and **2c** were similar to those for the corresponding per-O-acetyl derivatives **1b-1c**.

The EI mass spectrum of **3c** (see Experimental) shows as the main primary fragmentation the loss of acetic acid ( $m/z$  567). Peaks for the fluorenyl cation (165) and 2-amino-4,4-diphenyloxazoline were also observed.



## EXPERIMENTAL

General Procedures. Melting points were determined on a Gallenkamp MFB 595 apparatus and are uncorrected. Optical rotations were measured at 22 °C with a Perkin-Elmer 141 MC polarimeter. UV spectra were recorded using a Beckman DU-7 spectrophotometer. IR spectra for KBr discs were measured with a Perkin-Elmer 299 instrument. When solutions in carbon tetrachloride were used, the spectra were recorded on a Bomen Michelson MB-100 FT IR spectrophotometer; the solutions ( $1.00 \cdot 10^{-2}$ ,  $0.75 \cdot 10^{-2}$ ,  $0.50 \cdot 10^{-2}$  and  $0.25 \cdot 10^{-2}$  molar) were prepared with freshly distilled carbon tetrachloride and a fixed, 1.00 mm path length NaCl cell was used.  $^1\text{H}$  NMR (200.13 MHz) and  $^{13}\text{C}$  NMR (50.33 MHz) were obtained with a Varian XL-200 spectrometer for solutions in  $\text{CDCl}_3$  or  $\text{DMSO}-d_6$  with tetramethylsilane ( $\text{Me}_4\text{Si}$ ) as internal reference. The spectra are reported as chemical shifts downfield from  $\text{Me}_4\text{Si}$ . Assignments of  $^1\text{H}$  signals were confirmed by decoupling and H/D exchange experiments. Proton-decoupled APT $^{13}$  (attached proton test) spectra were used to assist in carbon signal assignments. Mass spectra (EI mode) were taken on a Kratos MS-80RFA instrument operated at an ionizing energy of 35 eV, ionizing current 100  $\mu\text{A}$ , accelerating voltage 4 KV, resolution 1000 (10% valley definition). The elemental composition of the ions was determined by a peak-matching method relative to PFK on the same instrument, the resolution being 10000 (10% valley definition). Metastable peaks in the field-free region were obtained on the same instrument; the fragment ion scan (FIS, B:E constant) was used. TLC was performed on silica gel 30 F $_{254}$  (Merck) plates with detection by UV light or/and by charring with 10% sulphuric acid, and column

chromatography was carried out with silica gel 60 (Merck). Microanalyses were performed in the Analytical Chemistry Department at Sevilla University.

N(N')-Hetaryl-N'(N)-(2,3,4,6-tetra-O-acetyl- $\beta$ -D-glucopyranosyl)-thioureas (1a-1d). A solution of the corresponding 2-aminoheterocycle<sup>8</sup> (1.28 mmol) in acetone (5 mL) was added dropwise to a solution of 2,3,4,6-tetra-O-acetyl- $\beta$ -D-glucopyranosyl isothiocyanate<sup>6,7</sup> (0.5 g, 1.28 mmol) in acetone (5 mL) under nitrogen. The resulting solution was refluxed (1a) or kept at room temperature for t h (1b-1d) and then concentrated under diminished pressure, and the residue was purified by column chromatography using an ether-hexane 4:1, 5:1, 6:1 gradient as eluant to give a white foam (1a, 1b, 1d) or solid crystallised from ethanol (1c).

N-(2,3,4,6-Tetra-O-acetyl- $\beta$ -D-glucopyranosyl)-N'-(thiazole-2-yl)-thiourea (1a, 0.113 g, 22%, t 4 h) had  $[\alpha]_D^{22}$   $-20.1^\circ$  (c 0.6, chloroform); UV (CHCl<sub>3</sub>) 296 and 253 nm ( $\epsilon_{mM}$  27.0 and 15.1); IR (KBr) 3470 and 3220 (NH), 1750 (C=O), 1620 (thiazole ring), 1550 (NH), 1505, 1425 (thiazole ring) and 1230 cm<sup>-1</sup> (C=S and C-O-C); <sup>1</sup>H NMR (CDCl<sub>3</sub>) Tables 1 and 2, and  $\delta$  2.09, 2.05, 2.03, and 2.02 (4s, 12H, 40Ac); <sup>13</sup>C NMR (CDCl<sub>3</sub>) Table 3 and  $\delta$  180.4 (C=S), 170.2, 169.8 (2C) and 169.3 (4C=O), 161.1 (C-2 heterocycle), 128.2 (C-4 heterocycle), 111.7 (C-5 heterocycle), 20.7 and 20.5 (3C) (4CH<sub>3</sub>); MS: m/z 331 (4, glycosidic moiety), 271 (2), 211 (3), 169 (11), 142 (80, thiazole-2-yl isothiocyanate), 127 (4), 109 (10), 100 (21, 2-aminothiazole), 85 (7), 73 (43) and 60 (100). Anal. Calcd for C<sub>18</sub>H<sub>23</sub>N<sub>3</sub>O<sub>9</sub>S<sub>2</sub>: C, 44.16; H, 4.74; N, 8.59. Found: C, 43.89; H, 5.00; N, 8.34.

N-(2,3,4,6-Tetra-O-acetyl- $\beta$ -D-glucopyranosyl)-N'-(2-thiazoline-2-yl)-thiourea (1b, 0.37 g, 58%, t 2 h) had  $[\alpha]_D^{22}$   $-3.9^\circ$  (c 1.0, chloroform);

UV ( $\text{CH}_2\text{Cl}_2$ ) 274 and 240 nm ( $\epsilon_{\text{mM}}$  12.4 and 9.0); IR (KBr) 3310 (N'H), 3080 (NH), 1745 (C=O), 1600 (C=N), 1550 (NH) and 1225  $\text{cm}^{-1}$  (C=S and C-O-C). (in  $\text{CCl}_4$ ) 3030 (NH) and 1609 (C=N);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ) Tables 1 and 2, and  $\delta$  2.08, 2.03, 2.02 and 2.01 (4s, 12H, 40Ac);  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ ) Table 3 and  $\delta$  182.8 (C=S), 170.6, 169.9, 169.5 and 169.4 (4C=O), 163.5 (C-2 heterocycle), 55.5 (C-4 heterocycle), 25.1 (C-5 heterocycle), 20.7, 20.6 and 20.5 (2C) ( $4\text{CH}_3$ ); MS:  $m/z$  331 (25, glycosidic moiety), 271 (4), 211 (4), 169 (68), 144 (11, 2-thiazoline-2-yl isothiocyanate), 127 (22), 109 (50), 102 (55, 2-amino-2-thiazoline) and 60 (100). Anal. Calcd for  $\text{C}_{18}\text{H}_{25}\text{N}_3\text{O}_9\text{S}_2$ : C, 43.98; H, 5.13; N, 8.55. Found: C, 43.92; H, 5.15; N, 8.47.

N-(4,4-Diphenyloxazoline-2-yl)-N'-(2,3,4,6-tetra-O-acetyl- $\beta$ -D-glucopyranosyl)thiourea (1c, 0.33 g, 40%,  $t$  24 h) had mp 97-98 °C;  $[\alpha]_{\text{D}}^{22} +9.1^\circ$  ( $c$  0.7 chloroform); UV ( $\text{CH}_2\text{Cl}_2$ ) 277, 251 and 234 nm ( $\epsilon_{\text{mM}}$  8.4, 21.8 and 11.5); IR (KBr) 3300 (NH), 1620 (C=N), 1600 (C=C aromatic), 1510 (NH), 1230 (C=S and C-O-C), 750 and 705  $\text{cm}^{-1}$  (CH aromatic);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ) Tables 1 and 2, and  $\delta$  7.45-7.20 (m, 10H, 2Ph), 2.08, 2.03 and 2.01 (3s, 12H, 40Ac);  $^1\text{H}$  NMR ( $\text{DMSO}-d_6$  at 130 °C) Table 1 and  $\delta$  7.50-7.20 (m, 10H, 2Ph), 2.02, 1.98, 1.96 and 1.92 (4s, 12H, 40Ac);  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ ) Table 3 and  $\delta$  193.3 (C=S), 170.6, 170.3, 169.8 and 169.4 (4C=O), 166.7 (C-2 heterocycle), 141.3-126.0 (12C, 2Ph), 79.7 (C-5 heterocycle), 77.1 (C-4 heterocycle), 20.6 and 20.4 (3C) ( $4\text{CH}_3$ ); MS:  $m/z$  331 (10, glycosidic moiety), 280 (2, 4,4-diphenyloxazoline-2-yl isothiocyanate), 280 (2), 271 (2), 238 (7, 2-amino-4,4-diphenyloxazoline), 211 (2), 169 (39), 165 (23, fluorenyl cation), 161 (100), 127 (12), 109 (27) and 60 (35). Anal. Calcd for  $\text{C}_{30}\text{H}_{33}\text{N}_3\text{O}_{10}\text{S}$ : C, 57.40; H, 5.30; N, 6.70. Found: C, 57.11; H, 5.23; N, 6.48.

N-(cis-3a,4,5,6,7,7a-Hexahydrobenzoxazole-2-yl)-N'-(2,3,4,6-tetra-O-acetyl- $\beta$ -D-glucopyranosyl)thiourea (1d, pair of diastereomers, 0.5 g, 73%, t 24 h) had UV ( $\text{CH}_2\text{Cl}_2$ ) 270 and 231 nm ( $\epsilon_{\text{mM}}$  10.8 and 4.7); IR (KBr) 3320 (NH), 3100 (N'H), 1750 (C=O), 1680 (C=N), 1560 (NH), 1230 (C=S and C-O-C), (in  $\text{CCl}_4$ ) 3038 (N'H) and 1684 (C=N);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ) Tables 1 and 2, and  $\delta$  2.75-1.10 (m, 8H,  $4\text{CH}_2$ ), 2.22, 2.20, 2.16 and 2.14 (4s, 12H, 4OAc);  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ ) major diastereomer Table 3 and  $\delta$  180.9 (C=S), 170.6, 169.9, 169.5 and 169.4 (4C=O), 155.3 (C-2 heterocycle), 75.5 (C-7a heterocycle), 58.5 (C-3a heterocycle), 26.3, 26.1, 21.3 and 19.3 ( $4\text{CH}_2$ ), 20.7 and 20.5 (3C) ( $4\text{CH}_3$ ), distinguishable signals of minor diastereomer table 3, and  $\delta$  180.6 (C=S), 156.5 (C-2 heterocycle), 75.8 (C-7a heterocycle), 58.7 (C-3a heterocycle), 26.2, 25.8, 21.2, 19.1 ( $4\text{CH}_2$ ), 20.5 (3C) and 20.3 ( $4\text{CH}_3$ ); MS:  $m/z$  331 (43, glycosidic moiety), 271 (7), 182 (4, hexahydrobenzoxazole-2-yl isothiocyanate), 169 (100), 140 (25, 2-aminohexahydrobenzoxazole), 127 (18), 109 (39) and 97 (61). Anal. Calcd for  $\text{C}_{22}\text{H}_{31}\text{N}_3\text{O}_{10}\text{S}$ : C, 49.90; H, 5.90; N, 7.94. Found: C, 49.74; H, 5.85; N, 7.89.

N(N')-Hetaryl-N'(N)-(2,3,4,6-tetra-O-benzoyl- $\beta$ -D-galactopyranosyl)-thioureas (2b, 2c). A solution of the corresponding 2-aminoheterocycle (0.785 mmol) in acetone (20 mL) was added dropwise to a solution of 2,3,4,6-tetra-O-benzoyl- $\beta$ -D-galactopyranosyl isothiocyanate<sup>7</sup> (0.5 g, 0.785 mmol) in acetone (5 mL) under nitrogen. The resulting solution was kept at room temperature for t h and then concentrated under diminished pressure, and the residue was crystallised from ethanol.

N-(2,3,4,6-Tetra-O-benzoyl- $\beta$ -D-galactopyranosyl)-N'-(2-thiazoline-2-yl)thiourea (2b, 0.4 g, 70%, t 1.5 h) had mp 124 °C;  $[\alpha]_{\text{D}}^{22} +150.4^\circ$  (c 1.0, chloroform); UV ( $\text{CH}_2\text{Cl}_2$ ) 233 nm ( $\epsilon_{\text{mM}}$  45.5); IR (KBr) 3320 (N'H),

3090 (NH), 1725 (C=O), 1600 (C=N and C=C aromatic), 1560 (NH), 1270 (C=S and C-O-C), 770 and 710  $\text{cm}^{-1}$  (CH aromatic), (in  $\text{CCl}_4$ ) 3098 (NH) and 1603 (C=N);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ) Tables 1 and 2 and  $\delta$  8.07–7.17 (m, 20H, 4Ph);  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ ) Table 3 and  $\delta$  182.7 (C=S), 166.2, 165.5 and 165.3 (2C) (4C=O), 162.7 (C-2 heterocycle), 133.3–128.1 (24C, 4Ph), 55.4 (C-4 heterocycle), 25.1 (C-5 heterocycle); MS:  $m/z$  579 (7, glycosidic moiety), 122 (29), 105 (100) and 77 (40). Anal. Calcd for  $\text{C}_{38}\text{H}_{33}\text{N}_3\text{O}_9\text{S}_2$ : C, 61.69; H, 4.50; N, 5.68. Found C, 61.90; H, 4.78; N, 5.42.

N-(4,4-Diphenyloxazoline-2-yl)-N'-(2,3,4,6-tetra-O-benzoyl- $\beta$ -D-galactopyranosyl)thiourea (2c, 0.43 g, 63%,  $t$  24 h) had mp 125 °C;  $[\alpha]_{\text{D}}^{22}$  +108.1° ( $c$  0.7, chloroform); UV ( $\text{CH}_2\text{Cl}_2$ ) 233 nm ( $\epsilon_{\text{mM}}$  57.1); IR (KBr) 3300 (NH), 1725 (C=O), 1620 (C=N), 1600 (C=C aromatic), 1270 (C=S and C-O-C), 750 and 710  $\text{cm}^{-1}$  (CH aromatic);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ) Tables 1 and 2, and  $\delta$  8.20–7.20 (m, 31H, 6Ph and N'H);  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ ) Table 3 and  $\delta$  191.9 (C=S), 166.3, 166.0 and 165.3 (2C) (4C=O), 163.5 (C-2 heterocycle), 141.3–126.0 (36C, 6Ph), 80.4 (C-5 heterocycle) and 77.2 (C-4 heterocycle); MS:  $m/z$  579 (6, glycosidic moiety), 2.80 (1, 4,4-diphenyloxazoline-2-yl isothiocyanate), 238 (9, 2-amino-4,4-diphenyloxazoline), 165 (20, fluorenyl cation), 161 (100), 105 (20) and 77 (9). Anal. Calcd for  $\text{C}_{50}\text{H}_{41}\text{N}_3\text{O}_{10}\text{S}$ : C, 68.56; H, 4.72; N, 4.80. Found: C, 68.67; H, 4.68; N, 4.60.

1,3,4,6-Tetra-O-acetyl-2-deoxy-2-[3-(4',4'-diphenyloxazoline-2'-yl)thioureido]- $\beta$ -D-glucopyranose (3c). A solution of 2-amino-4,4-diphenyloxazoline (0.29 g, 1.20 mmol) in acetone (15 mL) was added dropwise to a solution of 1,3,4,6-tetra-O-acetyl-2-deoxy-2-isothiocyanato  $\beta$ -D-glucopyranose<sup>23</sup> (0.46 g 1.20 mmol) in acetone (10 mL) under

nitrogen. The resulting solution was kept at room temperature for 24 h and then concentrated under diminished pressure, and the residue was purified by column chromatography using an ether-hexane 4:1, 5:1, 6:1 gradient as eluant. On recrystallisation from ethanol, white crystals of pure compound 3c were obtained (0.28 g, 37%), mp 103–104 °C;  $[\alpha]_D^{22}$  0° ( $c$  1, chloroform); UV ( $\text{CH}_2\text{Cl}_2$ ) 276, 256 and 230 nm ( $\epsilon_{\text{mM}}$  13.0, 23.6 and 7.8); IR (KBr) 3320 and 3170 (NH), 1750 (C=O), 1625 (C=N), 1600 (C=C aromatic), 1525 (NH), 1230 (C=S and C–O–C), 755 and 705  $\text{cm}^{-1}$  (CH aromatic);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ) Tables 1 and 2, and  $\delta$  7.38–7.24 (m, 10H, 2Ph), 2.10, 2.08, 2.04 and 2.00 (4s, 12H, 4OAc);  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ ) Table 3 and  $\delta$  191.1 (C=S), 170.6, 170.3, 169.2 and 169.0 (4C=O), 165.8 (C-2 heterocycle), 141.1–126.0 (12C, 2 Ph), 77.1 (in  $\text{DMSO}-d_6$ , C-5 heterocycle), 74.3 (in  $\text{DMSO}-d_6$ , C-4 heterocycle), 20.8, 20.7, 20.6 and 20.5 (4 $\text{CH}_3$ ); MS:  $m/z$  567 (32,  $\text{M}^+$ -AcOH), 537 (8), 508 (89), 507 (61), 490 (83), 448 (29), 405 (40), 363 (32), 238 (9, 2-amino-4,4-diphenyloxazoline), 222 (23), 180 (60), 179 (85), 165 (75, fluorenyl cation), 161 (71) and 77 (29). Anal. Calcd for  $\text{C}_{30}\text{H}_{33}\text{N}_3\text{O}_{10}\text{S}$ : C, 57.41; H, 5.30; N, 6.70. Found: C, 57.60; H, 5.52; N, 6.43.

#### ACKNOWLEDGMENTS

We thank the Ministry of Education and Science of Spain for the awards of scholarships (to J.M.G.F. and T.C.L.) and the D.G.I.C.Y.T. for financial support (grant PB 88-0268).

#### REFERENCES AND FOOTNOTES

1. Presented at the Fifth European Symposium on Carbohydrates, Prague, Czechoslovakia, August 21–25, 1989. Abst. Pap. A-113.

2. J. Witzack, Adv. Carbohydr. Chem. Biochem., **44**, 91 (1986) and references therein.
3. J. Fuentes Mota, J. M. García Fernández, C. Ortiz Mellet, M. A. Pradera Adrián and R. Babiano Caballero, Carbohydr. Res., **193**, 314 (1989) and references therein.
4. C. Gmernicka-Haftek and W. Wieniawski, Acta Pol. Pharm., **24**, 253 (1967).
5. M. Avalos González, R. Babiano Caballero, P. Cintas Moreno, J. Fuentes Mota, J. L. Jiménez Requejo and J. C. Palacios Albarrán, Heterocycles, **29**, 1 (1989) and Nucleosides and Nucleotides, **9**, 137 (1990).
6. F. P. Kamp and F. Micheel, Chem. Ber., **89**, 133 (1956).
7. R. Babiano Caballero, J. Fuentes Mota and J. A. Galbis Pérez, Carbohydr. Res., **154**, 280 (1986).
8. 2-Aminothiazole and 2-amino-2-thiazoline are commercial products; 2-amino-*cis*-3a,4,5,6,7,7a-hexahydrobenzoxazole was described by R. R. Wittekind, J. D. Rosenau and G. I. Poos, [J. Org. Chem., **26**, 444 (1961)] and 2-amino-4,4-diphenyloxazoline (mp 176 °C) was prepared from 1,1-diphenyl-2-iodoethylisocyanate [A. Hassner, M. E. Lorber and C. Heathcock, J. Org. Chem., **32**, 540 (1967)] by reaction with ammonium hydroxide and cyclodehydrohalogenation (J. Fuentes Mota, C. Ortiz Mellet, M. A. Pradera Adrián and J. M. García Fernández, unpublished results).
9. J. C. Grasselli, Ed.; Atlas of Spectral Data and Physical Constants for Organic Compounds, C.R.C. Press: Cleveland, 1972, p. B-930.
10. A. R. Katritzky and A. P. Ambler in Physical Methods in Heterocyclic Chemistry, Vol II; A. R. Katritzky, Ed.; Academic Press: New York, 1963, p. 218.
11. A. J. Castro, J. P. Marsh Jr. and B. T. Nakata, J. Org. Chem., **28**, 1943 (1963).
12. E. Prestch, T. Clerc, J. Seibl and W. Simon. Tablas para la elucidación estructural de compuestos orgánicos por métodos espectroscópicos, Alhambra: Madrid, 1980, p. H-80.
13. S. L. Patt and J. N. Shoolery; J. Mag. Reson., **46**, 535 (1982).
14. J. Fuentes Mota, J. M. García Fernández, C. Ortiz Mellet, M. A. Pradera Adrián and M. García Gómez, An. Quim., in press.
15. K. Bock and C. Pedersen, Adv. in Carbohydr. Chem. Biochem., **41**, 27 (1983).
16. W. Walter and K. P. Ruess, Liebigs Ann. Chem., **746**, 54 (1971).

17. M. L. Martín, M. L. Filleux-Blanchard, G. J. Martín and G. A. Webb, Organic Mag. Reson., **13**, 396 (1980).
18. M. Avalos, R. Babiano, P. Cintas, J. L. Jiménez, J. C. Palacios and J. Fuentes; J. Chem. Soc., Perkin Trans. I, 495 (1990) and references therein.
19. J. Fuentes Mota, M. C. Ortiz Mellet, F. Segura Ramos, M. A. Pradera Adrián and A. Cert Ventulá; An. Quim., **79C**, 221 (1983) and references therein.
20. G. B. Waller Ed.; Biochemical Applications of Mass Spectrometry, Wiley Interscience, New York, 1972, p. 335.
21. N. K. Kochetkov and O. S. Chizhov in Methods in Carbohydrate Chemistry, Vol VI; R. L. Wistler and J. N. Bemiller, Eds.; Academic Press: New York, 1972, p. 540.
22. Q. N. Porter; Mass Spectrometry of Heterocyclic Compounds, Wiley Interscience: New York, 1985, p. 894, 899 and 905.
23. J. C. Jochims and A. Seeliger, Tetrahedron, **21**, 2611 (1965).